# An Ancestral Oomycete Locus Contains Late Blight Avirulence Gene *Avr3a*, Encoding a Protein that is Recognised in the Host Cytoplasm

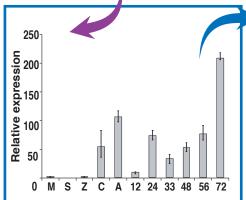
Miles R Armstrong<sup>1</sup>, Stephen C Whisson<sup>1</sup>, Leighton Pritchard<sup>1</sup>, Jorunn I B Bos<sup>2</sup>, Eduard Venter<sup>1</sup>, Anna O Avrova<sup>1</sup>, Anne P Rehmany<sup>3</sup>, Ulrike Böhme<sup>4</sup>, Karen Brooks<sup>4</sup>, Inna Cherevach<sup>4</sup>, Nancy Hamlin<sup>4</sup>, Brian White<sup>4</sup>, Audrey Fraser<sup>4</sup>, Angela Lord<sup>4</sup>, Michael A Quail<sup>4</sup> Carol Churcher<sup>4</sup>, Neil Hall<sup>4</sup>, Matthew Berriman<sup>4</sup>, Sanwen Huang<sup>5</sup>, Sophien Kamoun<sup>2</sup>, Jim L Beynon<sup>3</sup> and Paul R J Birch<sup>1</sup> *Scottish Crop Research Institute*; <sup>2</sup>The Ohio State University; <sup>3</sup> Warwick HRI; <sup>4</sup>The Wellcome Trust Sanger Institute; **Research Institute**; <sup>5</sup>Wageningen University

#### **INTRODUCTION:**

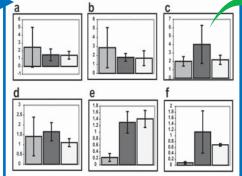
The oomycete *Phytophthora infestans* causes late blight, the potato disease that precipitated the Irish famines in 1846 and 1847. It represents a re-emerging threat to potato production and is one of over 70 species which are arguably the most devastating pathogens of dicotyledonous plants. Nevertheless, little is known about the molecular bases of pathogenicity in these algae-like organisms, or of avirulence molecules that are perceived by host defences. Disease resistance alleles, products of which recognise corresponding avirulence molecules in the pathogen, have been introgressed into the cultivated potato from a wild species, *Solanum demissum*, and *R1* and *R3a* have been identified. We used association genetics to identify *Avr3a*, and show that it encodes a protein that is recognised in the host cytoplasm, where it triggers *R3a*-dependent cell death (Armstrong *et al* (2005) *Proc. Natl. Acad. Sci* (USA) 102:7766-7771.

### **RESULTS:**

We used association Hvaloperonospora parasitica The Avr3a locus revealed coding sequences (CDSs) genetics to show that 9b13+4c4 1g5-18 12i13-17 12i13-18 Pex147 (encoding a similar to CDSs flanking ATR1NdWsB in 147 amino acid secreted >126kb 1.2kb Hyaloperonospora protein) was likely to be parasitica. MTD, a RAS-like Avr3a. Pex147 was PCR >10kb protein, F-actin capping amplified from 55 protein and 3-IPMDH were virulence-tested P. infestans. Three SNPs co-linear in the loci. ATR1NdWsB and Avr3a were found, changing amino acids S19C, E80K share little sequence similarity, but are in similar and M103L The SNPs 10.9kb +9.6kb relative locations. Within revealed two alleles these co-linear loci, the correlated 100 % with virulence phenotype on avirulence genes have evolved differently. In cv. Pentland Ace, H. parasitica, considerable containing R3a. The C19 allelic variation was K80 I103 (C-K-I) allele was 12 Ĥ, 1kb observed in ATR1NdWsB. associated with avirulence, 48kb 11kb 33kb 31kb H Ŵ X ÿ 24 In P. infestans, gene while virulent isolates were × duplication and divergence homozygous for the S19 124kb 1kb 42kh 9kh 2kb 0 5kb 4kb 26kb >12kb 0.2kh has generated variation in E80 M103 (S-E-M) allele. BACs containing Pex147 Avr3a-like sequences and Pi-BAC-49P21 were sequenced to reveal numerous synonymous and non-synonymous SNPs two paralogues of Pex147 Pi-BAC-61F2 61F2 Phytophthora infestans distinguish these genes. (Pex147-2 and Pex147-3)



Expression of *Pex147* was readily detectable in pre-infection (sporangia, zoospores, germinating cysts, and appressoria) and infection (susceptible potato cv. Bintje, 12, 24, 33, 48, 56 and 72 hpi) stages. *Pex147* was up-regulated >100-fold in appressoria and showed elevated levels of expression throughout infection, with an early peak of expression at 24 hpi, in the biotrophic phase of infection, and >200-fold elevation of expression at 72 hpi, in the necrotrophic phase of the interaction.



Truncated E-M (virulent) and K-I (avirulent) alleles of *Pex147* were co-expressed with a vector expressing gfp, as a marker of cell vitality, in potato genotypes lacking an *R* gene (cv. Bintje), or containing *R3a, R3b, R1, R2 and R10.* There was a 5-fold reduction in GFP fluorescence in Pentland Ace (R3a) co-bombarded with the truncated K-I sequence, indicating a reduction in vital cells consistent with triggering HR. This was also seen in transgenic Desireé expressing *R3a* and *Pex147* was renamed *Avr3a*.



Co-infiltration of *N. benthamiana* with *A. tumefaciens* carrying a construct expressing *R3a*, and a strain expressing the truncated K-I *Avr3a* sequence resulted in confluent cell death. In contrast, co-expression of *R3a* with the truncated E-M *avr3a* sequence, or infiltrations of individual *A. tumefaciens* strains and other controls, failed to elicit visible cell death.

### **CONCLUSION:**

Association genetics was used to identify the P. infestans Avr3a gene and show that its product is recognised in an R3a¬dependent manner in the host cytoplasm. Analysis of the Avr3a locus revealed unexpected conservation of synteny with the locus containing ATR1NdWsB in H. parasitica, supporting comparative genomics as an approach to investigate evolution of pathogenicity in oomycetes. The isolation of R3a and Avr3a represents an opportunity to investigate the earliest recognition events in a potato-P. infestans R-AVR interaction, and of subsequent signalling pathways leading to disease resistance. It also opens a door to studies of molecular mechanisms potentially underlying the biotrophic and necrotrophic phases of the P. infestans infection cycle.

## **ACKNOWLEDGEMENTS**

This work was funded by the Scottish Executive Environment and Rural Affairs Department