

The Role of Protease Cathepsin B during non-host HR

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Introduction

Programmed cell death (PCD) is a fundamental process in development and in responses to biotic and abiotic stresses. Caspase activities are involved in PCD in both animals and plants, although the genes encoding them are highly dissimilar. In addition, the cysteine protease cathepsin B has been demonstrated to act as both an effector and regulator in animal PCD, where it is implicated in relocalisation of cytochrome c from mitochondria to the cytosol. We demonstrate that cathepsin B is also required for PCD in a plant disease resistance hypersensitive response (HR). The bacterium *Erwinia amylovora* (*Eam*) induced a rapid non-host HR in *Nicotiana benthamiana*. The HR was preceded by a transient increase in cathepsin B activity and coincident early relocalisation of cytochrome C. These events and the subsequent HR were prevented by cathepsin B-specific inhibitors. Virus-induced gene silencing (VIGS) was used to confirm that cathepsin B was essential for *Eam*-mediated HR and showed that this HR was dependent on the ubiquitin ligase-associated protein SGT1. The requirement for cathepsin B in the HR reveals a structurally conserved cysteine protease in an ancient PCD mechanism shared by plants and animals.

Cathepsin B-specific inhibitors delay the HR

To investigate involvement of cathepsin B in the *Eam*-mediated HR, cathepsin B-specific inhibitors I (Z-FA.fmk), II (Ac-LVK-CHO), IV (CA-074 Me) and the broad cathepsin B, S, L and papain inhibitor Z-FGNHO-Bz were each infiltrated with 10⁶ cfu ml⁻¹ of *Eam* into *N. benthamiana* leaves. In the absence of inhibitor, *Eam* elicited a visible HR by 24 hours post-infiltration (hpi). In contrast, infiltration of *Eam* in the presence of 1 mM of inhibitors consistently abolished or considerably delayed and reduced the HR.

N. benthamiana leaves 4 dpi with 10° cfu/ml Eam +/- 1 mM inhibitors

CathB Inhibitor Perturbs Cyt. C Release from Mitochondria

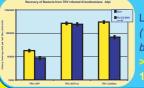
Western blots indicating presence of cytochrome c or NAD9 proteins in cytoplasmic and mitochondrial cellular fractions prepared from untreated leaves (U); leaves 6, 12 or 18 hpi with *Eam*; *Eam* with 1 mM Z-FA.fmk (Cathepsin B I) inhibitor. The same result was observed using 1 mM Ac-LVK-CHO (Cathepsin B II) inhibitor. Equal loading is indicated by Ponceau staining.



Cathepsin B-specific inhibitor prevents Eam-induced relocalisation of cytochrome c from the mitochondria to the cytoplasm.

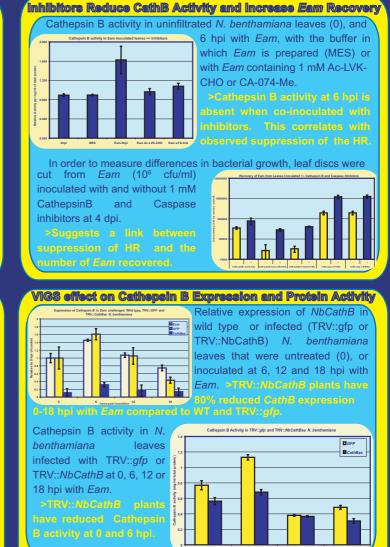


N. benthamiana inoculated with TRV::gfp, TRV::NbCathB and TRV::Nbsgt1 were infiltrated with 10⁵ or 10⁶ cfu.ml⁻¹ Eam. >VIGS of NbCathB suppresses the HR



Leaf discs were cut from *Eam* and *Pst* (10⁶ cfu/cm²) inoculated VIGSed *N. benthamiana.*

0-fold greater bacterial growth 4 dpi.



Conclusion

We propose that PCD in plants, as in animals, involves a cascade of interacting proteases, including cysteine proteases. Although caspase activities are involved in both plant and animal PCD, the genes encoding them are not similar. In contrast, the requirement for cathepsin B in the HR has revealed an ancient PCD mechanism executed or regulated by a cysteine protease that is structurally conserved in plants and animals.

We are grateful to SEERAD and BBSRC for funding this work. We thank Dr J-M. Grienenberger (IBMP-CNRS, Strasbourg, France) and Dr K. Shirasu (Sainsbury laboratory, Norwich, UK) for, respectively, providing anti-NAD9 and anti-SGT1 antisera.