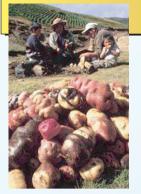
Carotenoid accumulation during potato tuber development and storage



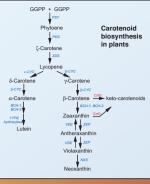


W L Morris, L Ducreux, D W Griffiths, D Stewart, H V Davies and M A Taylor Quality, Health & Nutrition, Scottish Crop Research Institute, Invergowrie, Dundee, DD2 5DA.

Introduction

The health benefits associated with the availability of carotenoids in the human diet are becoming increasingly apparent. Carotenoids cannot be synthesised by vertebrates and therefore need to be provided via dietary intake. There would be an undoubted benefit to health therefore if the carotenoid content and balance could be improved in a staple food such as notato

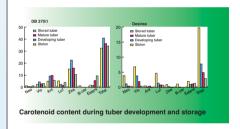
Tuber flesh colour can range from white through to orange and is a direct consequence of the presence of carotenoids. The molecular basis for this wide variation in tuber carotenoid content is not understood. The aim of this study is to compare carotenogenesis in white, yellow and orange-fleshed potato germplasm in order to reveal factors affecting carotenoid levels. This knowledge will then be used to identify possible targets for a transgenic approach.



Results

Comparison of carotenoid profiles during tuber development and storage

The levels of carotenoids during tuber development and storage were compared in a high carotenoid accumulating S. phureja accession (DB375\1) with two S tuberosum cultivars (Pentland Javelin and Desiree) that accumulate lower levels of tuber carotenoid. In S. phureia tubers at maturity the major carotenoids were zeaxanthin, antheraxanthin and violaxanthin. Following 9 months storage at 4°C the levels of zeaxanthin and antheraxanthin decreased whereas the level of lutein increased however. overall there was only a small decrease in total carotenoid content.



Creation of constructs for plant

Partial cDNAs of the genes involved in carotenoid

biosynthesis were cloned from S. phureia DB375\1

tuber cDNA by RT-PCR using primers designed to

conserved regions of publicly available sequences.

expression studies. Lycopene-E-cyclase (E-cyc) and

lycopene- β -cyclase (β -cyc) clones were used in the

These fragments were used as probes in gene

assembly of antisense constructs. An algal

(Haematococcus pluvialis) crtO over-expresser

construct was also produced. The constructs were

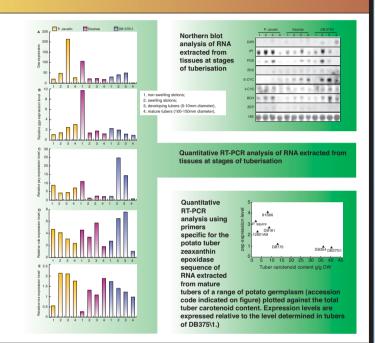
driven by the patatin promoter in order to confine

Constructs used in Agrobacterium-mediated potato

transformation

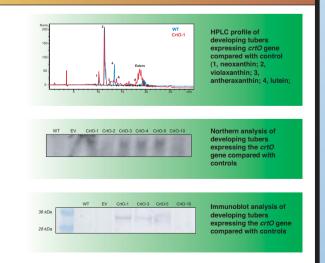
Biosynthetic gene expression profiling

The transcript levels of the genes encoding carotenogenic enzymes have been profiled in a range of germplasm during tuber development. Significant differences in the expression profiles were detected; suggesting that transcriptional control or mRNA stability gives rise to the large variation in tuber carotenoid content. Surprisingly, an inverse relationship between zeaxanthin epoxidase transcript level and total tuber carotenoid content was detected



Transgenic modification of tuber carotenoid content

Constructs have been assembled to either down-regulate potato genes encoding the carotenogenic enzymes (β -cyc and ϵ -cyc) or to over-express the algal crtO gene. Analysis of the effects of these transgenes is currently underway. Results are shown for the effects of over-expressing the algal *crtO* cDNA, encoding β -carotene ketolase in S. tuberosum cv Desiree. Carotenoid profiling reveals significant changes in tuber carotenoid levels. Transgenic plants contain lower levels of lutein and higher levels of carotenoid esters. More importantly, perhaps, is the presence of the commercially and nutritionally important keto-carotenoid astaxanthin in the transgenic tubers.



Work in Progress

the effects to the tuber.

-0-6

-FH

transformation

Acknowledgements

Transgene stacking experiments to enhance tuber carotenoid manipulation. Detailed analysis of transgenic potato lines with modified carotenoid content. Microarray experiments using oligonucleotide arrays of carotenoid biosynthetic genes.

ЪП

We gratefully acknowledge the help and advice of Professor A Young, and Dr D. Phillip, John Moores University, Liverpool for assistance in measuring cardenoids. We also thank Dr Norhiko Misawa, Kirin Brewery Co. Ltd, for the plasmid pACCRT-EIB and Prof. Joseph Hirschberg for the plasmid pPTCRTOBIB. This work was funded by the Scottish Executive Environment and Rural Affairs Department.