Screening wild potato accessions for resistance to the virulent allele of the Phytophthora infestans effector avr3a EM

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The perception of the Phytophthora infestans avirulence gene product Avr3a by the cognate potato resistance gene R3a results in a localised form of programmed cell death termed the hypersensitive response (HR), which inhibits pathogen development and mediates resistance (1). Pathogen effectors such as Avr3a, if not recognized by the plant, are thought to play a positive role during infection and in mediating susceptibility, and are therefore maintained by the pathogen. To prevent recognition, the pathogen is under constant selective pressure to alter the form of these effectors. Screening of P. infestans isolates has identified two alleles from Avr3a that display 100% correlation with either avirulence (Avr3a with the amino acids C¹⁹ K⁸⁰ and I¹⁰³) or virulence (avr3a with the amino acids S¹⁹ E⁸⁰ and M¹⁰³) in potato plants harbouring R3a. Our aim was to screen wild potato accessions, resistant to P. infestans, that recognise the mature virulent allele from Avr3a (avr3a EM) and/or the mature avirulent allele (Avr3a KI).



Screening the Commonwealth Potato Collection to identify potato accessions that recognise avr3a (EM)

We have screened 52 potato accessions from the Commonwealth Potato Collection (CPC), which have been identified as resistant or very resistant to P. infestans, by overexpressing avr3a EM and Avr3a KI utilising Agrobacterium tumefaciens-Potato Virus X (PVX) or A. tumefaciens only. In total, 22 accessions were responsive, of which 14 recognised both the virulent and avirulent alleles, 7 the virulent allele only and one the avirulent form only. The most consistent results were obtained with Solanum verrucosum, S. stoloniferum and S. demissum (Table 1). Co-bombardment of the Avr3a alleles with GFP has been used to confirm the recognition of avr3a EM and Avr3a KI in S. stoloniferum and S. microdontum (Figure 2). R3a-like sequences have been PCR amplified from the diploid Solanum species, S. verrucosum and are currently being analysed (Figure 3).

GFP & avr3a (EM)

	-	Species	Country	Series	EM	Accessions
-•	•	S commersonii	Arg/Bra/Uru	Commersoniana	1.1	5858 (1:3)
	5x	S semidemissum	Mex	Demissa	1:1	7103 (3:5)
	6v	S.brachycarpum	Mex	Demissa	0:1	1100 (0.0)
	6.	S. domingum	Mox	Domissa	1.11	2008 (4.0)
	6	S.uemissum S.ionatalum	Max	Demissa	1.11	2078 (4:7)
	ox	S.lopelalam	IVICX	Demissa	1.1	7055 (2:5)
	-	S.brevidens	Arg	Etuberosa	1:1	2451 (3:9)
	4x	S.fendleri	Mex/USA	Longipedicellata	1:2	4020 (3:7)
	4x	S.hjertingii	Mex	Longipedicellata	1:1	3029 (2:2)
	4x	S.papita	Mex	Longipedicellata	1:2	7085 (2:6)
	4x	S.polytrichon	Mex	Longipedicellata	1:2	3984 (2:4)
	4x	S.stoloniferum	Mex	Longipedicellata	4:10	2619 (4:9), 4013 (2:6), 2711 (3:4), 2220 (5:5), 1331 (2:2)
		S.cardiophyllum	Mex	Pinnatisecta	0:1	
		S.polyadenium	Mex	Polyadenia	0:1	
		S.alandiae	Arg/Bol/Chi	Tuberosa	0:1	
		S.berthaultii	Bol	Tuberosa	1:1	5701(1:4)
	-	S.microdontum	Arg	Tuberosa	3:7	4048 (1:3) , 7175 (3:6), 7176 (2:3)
	-	S.okadae	Arg	Tuberosa	0:1	
		S.venturii	Arg	Tuberosa	0:1	7327 (1:4)
	2x	S.vernei	Arg	Tuberosa	0:1	
		S.verrucosum	Mex	Tuberosa	2:2	7091(1:2), 7213 (3:5)
		S.chacoense	Chi/Arg/Par	Yungasensia	1:4	7211(2:5)
	/	Σ 21	8	8	21:52	14x EM&KI, 7x EM, 1x KI



N. benthamiana





S.Papita 7085



S. Hjt 3029

S. Ver 7213

Figure 1. A) Transient expression and coexpression of Avr3a and R3a in N. benthamiana. Leaves were infiltrated with A. tumefaciens carrying pGR106::Avr3a KI (or EM) alone, or mixed with a A. tumefaciens strain carrying pBIN plus::R3a. Similar results were obtained with a PVX free overexpression system utilising a pGRAB::Avr3a KI (or EM) based delivery (results not shown). B: PVX resistance response in Solanum papita. Recognition of avr3a Em and Avr3a KI in S. hjertingii (C) and S. verrucosum (D). Expression of the red fluorescent protein tdTomato (tdT) was used as a control

Table 1. A selection of 52 potato accessions comprising 21 potato species from 8 countries and 8 series, all resistant to P. infestans race 1,2,3,4,6,7 were screened and analysed for their response to overexpression of Avr3a KI and avr3a EM. The number of accessions responding to one or both alleles is shown in column 4. The CPC accession identifier and the number of independent, positive responses are shown in column 5. Accessions responding to avr3a EM are shown in BLUE, those responding to Avr3a KI shown in RED and those responding to both alleles in GREEN

GFP & Avr3a (KI) 4.00 3.00 2.50 3.00 2.00 2.00 1.50 1.00 1.00 0.50 CR STO 0.00 0.00

Figure 2. Preliminary co-bombardment results confirm recognition of avr3a EM and Avr3a KI in S. stoloniferum and S. microdontum but only recognition of Avr3a KI in S. tuberosum cv. Pentland Ace (R3a). Avr3a alleles were transiently coexpressed with GFP in different Solanum accessions, each compared with cv. Craigs Royal. In each histogram, the average number of GFP fluorescence cells is shown. This number was obtained by analysing 10 individual observation points (approximately 25 epidermal cells per point) for each bombarded leaf half. Co-bombardment with GFP and GUS resulted in comparable levels of GFP expressing cells in all potato accessions (results not shown).

Literature cited

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² Bos, J.I. et al. (2006) The C-terminal half of Phytophthora infestans RXLR effector AVR3a is sufficient to trigger R3a-mediated hypersensitivity and suppress INF1-induced cell death in Nicotiana benthamiana. Plant J. 48:165-176.

³ Huang, S. et al (2005) Comparative genomics enabled the isolation of the R3a late blight resistance gene in potato. Plant J, 42:251-261.



Figure 3. Restriction enzyme digest (Hhal) of R3a-like gene fragments from S. verrucosum. As the mature alleles of Avr3a differ only in two amino acids and R3a, which recognises Avr3a KI, has also been shown to weakly recognise avr3a EM (2) we used a PCR approach to amplify R3a-like sequences from S. verrucosum, which recognises both Avr3a alleles. Conserved primers, derived from the four known R3a paralogues (3), amplified a fragment of the expected size (3 kb) from genomic DNA comprising the ATG start codon at the 5'end but not the entire 3'UTR. A set of 77 cloned fragments were successfully digested and clustered into 17 groups according to their restriction enzyme pattern. Two main groups were identified (I) and (II) comprising 37 and 14 clones respectively. Group III comprised 5 clones. The remaining groups comprised between 1 and 3 members only. Sequencing of members from the different groups is currently in progress.